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Case Report

Case report of saccharomyces cerevisiae infection: root-cause analysis and remediation study

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Abstract

Introduction: Saccharomyces cerevisiae, used as yeast in producing foods such as beer, wine, and bread, is present in human microbiota and the natural environment. However, the increase in infection cases in recent years is remarkable.

Case Presentation: This report presents a urinary tract infection caused by S. cerevisiae in a patient with no history of probiotic use. The root-cause analysis of the infection and the improvement study were developed accordingly. In this context, it was found that the healthcare personnel who inserted a urinary catheter into the patient during hospitalisation in the intensive care unit fermented wine at home and checked the wine every morning. However, it could not be confirmed whether the isolated strain was the same as the strain in the urine culture since molecular-level studies could not be performed. Failure to comply with infection control measures during the treatment and care of at-risk individuals can lead to S. cerevisiae infection, an opportunistic pathogen.

Conclusion: By identifying the source of infection through root-cause analysis and carrying out improvement activities to the institution's characteristics, the healthcare professional's awareness can be raised, compliance with infection control measures can be increased, and behavioural change can be achieved.

Keywords

aseptic technique, hand hygiene, remediation study, root cause analysis, saccharomyces cerevisiae

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Introduction

S. cerevisiae, known as yeast, used in producing foods such as beer, wine, and bread, is present in human microbiota and the environment [1]. It is generally non-pathogenic [1,2]. The use of S. boulardii, a subtype of S. cerevisiae, as a probiotic in treating diarrhoea due to antibiotic use in recent years has increased infection [1]. Risk factors such as intensive care unit (ICU) hospitalisation, high-risk patient groups (such as leukaemia), elderly, premature children, corticosteroid, antibiotic, and probiotic use, central venous catheters (CVC), immunosuppression, hand hygiene non-compliance are predisposing factors for invasive Saccharomyces infections [2]. In recent years, Saccharomyces infections have increased due to increased immunosuppressive patients and developments in molecular diagnostic methods. Cases of fungemia, prosthetic valve endocarditis, peritonitis, cholecystitis, pneumonia, empyema, liver abscesses, vaginitis, and urinary tract infection (UTI) indistinguishable from Candida albicans have been reported in the literature [1,3,4]. This report presents a case of urinary tract infection (UTI) caused by S. cerevisiae in a patient with no history of probiotic use, the root-cause analysis, and the improvement study developed with the FOCUS-PDCA model. This case report has been prepared in accordance with the CARE case report guide.

Case Presentation

A 62-year-old woman with diabetes, hyperthyroidism, and right heart failure was admitted to the emergency department with complaints of chills, fever, malaise, abdominal pain, and dyspnoea. The patient did not smoke or drink alcohol. Blood pressure was 100/60 mmHg, heart rate 88/min, temperature 39.4° C, SPO2 88. Physical examination revealed (++) pretibial oedema, oliguria, dysuria, partial rales based on lung auscultation, and orthopneic dyspnoea. No suspicious findings related to infection were found on chest radiography. Laboratory examination revealed proteinuria, haematuria, bacteriuria, pancytopenia, neutropenia, leucocytosis, increased C-reactive protein (CRP), and procalcitonin levels. The patient was hospitalised in the infectious diseases ward, and treatment and follow-up were started. Vancomycin 2x1, meropenem 3x1, and enoxaparin sodium 1x0.4 IU were started empirically in addition to the drugs used due to chronic diseases. Filgrastim 1x48 IU (5 days) was added to the treatment with the recommendation of haematology for neutropenia. No growth was detected in blood and urine cultures taken on the first day of hospitalisation in the infectious diseases ward. On the third day of hospitalisation, renal function started to deteriorate. The dose of vancomycin and meropenem was changed to 1x1 g according to creatinine clearance. The patient was transferred to the Internal Medicine Intensive Care Unit (ICU) on the fourth day. In the ICU, a CVC was placed in the right femoral vein, and haemodialysis with ultrafiltration was performed for 4 hours for three consecutive days. A urinary catheter (UIC) was also inserted. On admission to the ICU, the patient had sub-febrile fever, hypotension, tachycardia, and macroscopic haematuria. No growth was detected in blood and urine cultures taken consecutively on the first day of ICU admission. On the eighth

Table 1. Urine culture results

Medicines	*S. cerevisiae MIC (µg/mL)
Voriconazole	4
Fluconazole	0.06
Amphotericin B	0.25
Caspofungin	16

*4 days after urinary catheter insertion

Table 2. Laboratory findings of the patient during follow-up and treatment

	Day 1	Day 3	Day 4	Day 8*	Day 10	Day 16
Leukocyte (103 /µl)	190.390	17.620	15.580	4.980	39.400	66.460
Haemoglobin (g/dl)	9.5	9.4	9.3	9.4	9.7	9.9
Neutrophils (103 /µl)	1.032	2.146	2.497	2.650	32.030	56.470
Platelet (103 /µl)	98 000	102 000	129 000	320 000	312 000	503 000
Urea (mg/dL)	82	102	112	79	104	86
Creatinine (mg/dL)	3.2	3.9	4.1	2.38	1.7	1.9
Glomerular filtration rate ml/min/1.73 m²	27	13	12	21	32	55
CRP (mg/L)	201	196	243	187	99	77
LDH (U/L)	435	397	418	629	574	217
AST (U/L)	67	58	64	96	84	73
Procalcitonin ng/ml	1.3	0.9	0.7	3.6	1.85	1.5
Urine Analysis						
Haematuria	Positive	Positive	Positive	Positive	Positive	Negative
Proteinuria	Positive	Positive	Positive	Positive	Negative	Negative
Bacteriuria	Positive	Positive	Positive	Positive	Positive	Positive
Leukocyte	34	12	7	8	14	5
Bacteria	1	1	1	3	3	2
Yeast cell	1	1	3	228	2544	62
Squamous epithelium	85	32	27	3	6	7

*4 days after urinary catheter insertion,
CRP: C-reactive protein, LDH: Lactate dehydrogenase, AST: Aspartate aminotransfer

Table 3. Improvement study according to the FOCUS-PDCA model

STAGE	DEFINITION	PERIOD
FIND (To find)	Find a process (topic) to improve	- Infection control measures, healthcare-associated infections
ORGANISED (Organizing)	Organise a team that knows the processes	- Specialist in Infectious Disease and Clinical Microbiolog
		-Infection Control Nurse
		- Internal Medicine Intensive Care Unit (ICU) Physician in charge
		- ICU Nurse in charge
CLARIFY (to explain)	Explain/clarify existing knowledge of the process	- S. cerevisiae for the first time in our hospital (isolated in urine culture)
		-Non-compliance with hand hygiene and aseptic techniques
UNDERSTAND (To understand)	Understand the operations and variations within the selected process	- Root cause analysis was carried out using the fishbone model. It was found that the consultant in charge of ICU was fermenting wine at home and did not comply with hand hygiene rules.
		-It was found that aseptic techniques were not adequately followed when inserting a urinary catheter; the bladder was placed on the bed. At the same time, the patient was being cared for, preventing drainage, and there was no sterile green drape material.
		-S. cerevisiae was isolated. However, there was no confirmation that it was the same strain, as molecular testing was impossible.
SELECT (To choose)	Identify potential actions for improvement and determine process improvement	-Hand hygiene
		-Insertion of urinary catheter under aseptic conditions
		-Patient care with urinary catheter
		-Use of protective equipment
PLAN (Plan)	Plan what to do in improvement	-Providing theoretical and practical training
		-The infection control nurse should increase the frequency of observation
		-Participatory observation by the infection control nurse
		-Using a urinary catheter related infection prevention bundle (while being fitted with a urinary catheter)
		-Feedback
DO (To do)	Implement plans for improvement	-The training sessions were repeated for a month with the participation of all the health care professionals.
		-There has been a 50% increase in observation time in the ICU.
		-Participatory observation of urinary catheter insertion and patient care has been implemented.
		-A roster of ICU staff was created to enable all staff to be observed during working hours.
		-The infection control nurse or the ICU nurse completed the urinary catheter-related infection prevention bundle during working hours. It was completed by the nurse responsible for the patient out of working hours.
CHECK (Check)	Check the results on improvement	-Sterile green cover was provided, the team was informed, and its use was ensured.
		-The hand hygiene monitoring report found a 32% increase compared to the previous and next three months.
		-The urinary catheter-related infection prevention bundle is used regularly, with a 14% increase in compliance in the first three months.
		-During patient care, the catheter is clamped when the bladder is level with the patient.
		-Aseptic precautions are taken during the insertion of a urinary catheter, but hand hygiene is not carried out as desired before the insertion of the catheter.
ACT (Behaviors)	Take action to protect gains and drive improvements	-Healthcare professionals (residents, students) who change regularly are trained according to the planning phase, regardless of the time involved.
		-On the first working day in the ICU, training is provided to replace residents, nurses, and support staff during the planning phase.
		-Besides the compulsory training, all health professionals working in the ICU receive regular training in line with the planning phase.
		-A decision has been made to use the urinary catheter infection prevention bundle for one year.
		-It was decided that the infection control nurse would be part of the observational study for one year.

day of hospitalisation (4th day in the ICU), urine microscopy revealed an intense amount of yeast cells. Simultaneously, urine culture was incubated in Yeast Extract Glucose Chloramphenicol Agar FIL-IDF medium at 37° C for 24 hours. At the end of incubation, yeasts were detected in the colonies grown on the medium, and the isolates were identified as S. cerevisiae by the BD PhoenixTM Yeast ID system. According to the results of urine microscopy, the patient's urinary catheter (UC) was excluded, and fluconazole 1x100 mg (5 days) was added to the treatment. No growth was detected in the blood culture. Renal functions and vital signs started to improve. After the 10th day of hospitalisation, CRP and procalcitonin values began to improve, but leukocyte and neutrophil levels were found to increase continuously. Bone marrow biopsy was performed, and amyloidosis and multiple myeloma were considered. Myeloid young cells were predominantly detected in the peripheral smear. On the 12th day of hospitalisation, the patient was diagnosed with chronic myeloid leukaemia and transferred to the haematology clinic. No growth was detected in the urine culture sent for the control on the 16th day of hospitalisation. The patient was discharged on the 44th day of hospitalisation. The patient's culture results are shown in Table 1, and laboratory findings are shown in Table 2. The infection control physician and nurse performed a root cause analysis due to the growth in urine culture. Accordingly, it was determined that the patient did not use beer, wine, and probiotics and did not ferment bread at home, but a staff member working in the intensive care unit made wine at home. It was determined that the UIC was implanted by the staff fermenting wine during the patient's ICU admission. S. cerevisiae was detected in the wine sample. However, it could not be confirmed whether the isolated strain was the same as the strain in the urine

culture since it was impossible to perform molecular-level studies. With the decision of the infection control committee, improvement work was initiated with the FOCUS-PDCA model to prevent healthcare-associated infections and to increase the awareness of the multidisciplinary team (Table 3). In this context, all personnel working in the ICU were given repeated training on hand hygiene, use of protective equipment, insertion of UIC under aseptic conditions, and care of patients with UIC. The observation period of the infection control nurse was increased by 50%, and participant observation was provided as a role model. Missing personal protective equipment was provided. The ICU team was motivated to use the bundle to prevent UIC-associated infections. As a result, hand hygiene compliance increased by 32% in the first three months. The use of the UIC-associated infection prevention bundle increased by 14% and continued to be used for one year. Patient care with UIC was started to be performed appropriately. Compliance with aseptic techniques during UIC insertion was achieved, but hand hygiene compliance during catheter insertion still needed to be increased.

Discussion

S. cerevisiae can be colonised in the hospital when hand hygiene is inadequate [5] and cleaning and disinfection procedures are not done properly [1]. Live S. cerevisiae cells can be airborne up to one meter and remain on all living/non-living surfaces for approximately two hours [6,7]. Cassone et al. (2003) reported that three patients developed fungemia in the ICU due to insufficient attention to infection procedures, while probiotic treatment was administered to other patients [6]. Elkhihal et al. (2015) reported that they separated S. cerevisiae in the urine culture of a 40-year-old type 2 diabetic patient without probiotic use due to digestive system translocation [8]. Intense yeast was observed in the urine sample four days after UIC insertion and S. cerevisiae was isolated in urine culture. Our patient, who had no history of probiotic use, did not use probiotics to treat other patients in the ICU. As a result of the root-cause analysis, it was determined that the personnel who put UIC on the patient fermented wine at home and checked the wine every morning. The fact that S. cerevisiae can remain on bare hands even after hand washing [7] suggests that the most probable cause was insufficient hand hygiene and compliance with aseptic techniques during UIC insertion. One of the responsibilities of the infection control physician and nurse assigned to the infection control committee within the scope of the Infection Control Regulation of inpatient treatment institutions is to determine the cause of infection and carry out studies to solve and improve the problem. In this case, the possible causes of infection were discussed using the fishbone model, and improvement work was carried out with the FOCUS-PDCA model to solve the problem. In this context, the awareness of healthcare personnel and compliance with infection control measures were increased and the healthcareassociated infection rate decreased. Accordingly, no infection caused by S. cerevisiae has been encountered in our hospital for the last three years. Studies have reported that S. cerevisiae is sensitive to antifungal agents such as voriconazole, fluconazole, amphotericin B, caspofungin, micafungin [1,2,4]. It is seen that many cases in the literature were treated with fluconazole (5-15 days) [2,5,6,8]. However, cases are treated with caspofungin [3], amphotericin B, and a combination of micafungin and fluconazole [1]. In S. cerevisiae infection, fluconazole used for 5-15 days may be useful alone in medical treatment. In addition to medical treatment, an important strategy is removing the invasive device associated with the infection and terminating probiotics. In cases in the literature, it is reported that invasive devices such as CVC and UC are removed immediately after the development of infection [1,3,4,6]. It is even suggested that success can be achieved only by removing the invasive device [6]. Similar to the treatment and care strategies in the literature, our patient's UC was released, fluconazole treatment was applied, and success was achieved. In studies, age, high-risk patients (such as leukaemia, renal failure, diabetes), chemotherapy, radiotherapy, use of immunosuppressive agents, antibiotics, probiotics, ICU hospitalisation, CVC, UC, total parenteral nutrition, failure of healthcare personnel to provide proper hand hygiene, failure to comply with procedures for infection control measures have been reported as risk factors for S. cerevisiae infection [1-5]. In our case, in addition to chronic diseases such as diabetes, hyperthyroidism, and heart failure, the patient's diagnosis of leukaemia, the use of broad-spectrum antibiotics, the continuation of treatment and care in the ICU, and the use of UIC constitute risk factors for infection. However, transporting the microorganism to our hospital with contaminated hands and failing to ensure proper hand hygiene and aseptic techniques during UIC insertion constitute important preventable risk factors for infection.

Limitations

This study has several limitations. First, as a single-case report, the findings cannot be generalised to broader patient populations or different clinical settings. Second, although the possible source of S. cerevisiae contamination was identified through a root-cause analysis, molecular typing could not be performed; therefore, the genetic similarity between the isolate detected in the patient's urine culture and the isolate obtained from the wine sample could not be confirmed. Third, environmental sampling in the intensive care unit was not conducted, which limited the ability to comprehensively assess potential environmental reservoirs.

Conclusion

Non-compliance with infection control measures during treating and caring for individuals at risk of infection may cause S. cerevisiae infection, an opportunistic pathogen. By determining the source of infection through root-cause analysis and carrying out improvement activities by the institution's characteristics, the healthcare professional's awareness can be raised, compliance with infection control measures can be increased, and behavioural change can be achieved. In this direction, S. cerevisiae infection can be prevented, and the healthcare-associated infection rate can be reduced.

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Scientific Responsibility Statement
The authors declare that they are responsible for the article's scientific content, including study design, data collection, analysis and interpretation, writing, and some of the main line, or all of the preparation and scientific review of the contents, and approval of the final version of the article.

Animal and Human Rights Statement
All procedures performed in this study were in accordance with the ethical standards of the institutional and/or national research committee and with the 1964 Helsinki Declaration and its later amendments or comparable ethical standards.

Data Availability Statement
The datasets used and/or analyzed during the current study are not publicly available due to patient privacy reasons but are available from the corresponding author on reasonable request.

Conflict of interest
The authors declare that there is no conflict of interest.

Ethics Declarations
This study was approved by the Ethics Committee of Kayseri University (Date: 2019-06-28, No: 16)

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